Glycoconjugate J (1985) 2:105--108

Preliminary Communication

Selective Synthesis of Cerebrosides: (2S, 3R, 4E)-1-O-β-D-galactopyranosyl-N-(2'R and 2'S)-2²hydroxy**tetracosanoyl-sphingenine***

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Received March 23, **1985.**

Key words:.cerebroside, synthesis, 1H-NMR

The cerebrosides were first isolated by Thudicum in 1874 and the structures were established *byCarteretal,* in 1950 (for review, see [2]). In 1961 Shapiro and Flowers [3] reported the first total synthesis of a cerebroside 1 (Fig. 1) which was identified with the natural sample, onlythrough comparison of their i.r. data. In order to confirm the absolute configuration at C-2' of natural cerebroside 1, we describe here an unambiguous synthesis of two stereoisomeric cerebrosides 1 and 2, and found that the ${}^{1}H$ -NMR spectra of the synthetic 1 (Fig. 2) was completely identical with that of the natural cerebroside reported recently by Dabrowski *et al. [4].*

In planning the synthetic route, the target structures 1 and 2 were disconnected at the dotted lines to give three key synthetic intermediates 3, 4 and 5 or 6 (Fig. 1).

Abbreviations: Bu, butyl; Ph, phenyl; t-BuPh₂SiCI, t-butyldiphenylsilyl chloride; MTPA, a-methoxy-a-trifluoromethylphenylacetic acid; THF, tetrahydrofuran.

*Part 36 in the series "Synthetic Studies on Cell-surface Glycans", for part 35, see [1].

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Figure 1. Key intermediates. Abbreviations: EE, ethoxyethyl; SitBuPh₂, t-butyldiphenylsilyl.

Figure 2. 400 MHz ¹H-NMR of (a) synthetic (2'R)-cerebroside 1, and (b) synthetic (2'S)-cerebroside 2. The spectra were recorded at 65°C in ²H₆-dimethylsulfoxide/²H₂O, 49/1 by vol, for the sample after exchanging three times with ²H₂O. Values of δ_H are expressed in ppm downward from internal standard tetramethylsilane. Values of $\frac{3}{H}$ _{HH} (Hz) are described in parentheses.

Figure 3. Synthetic scheme. Abbreviations: Bz, benzoyl; MTPA, a-methoxy-a-trifluoromethylphenylacetic acid.

The D-galactosyl donor 3 [5] and the (2S, 3R, 4E)-sphingenine derivative 4 [6] are already reported. The protected hydroxyacids 5 and 6 are readily prepared as follows (Fig. 3). Ethyl (2S)-2-acetoxy tetracosanoate 7, $\alpha|_D$ -14.2° (c 1.58, CHCl₃), obtainable [7] from (S)-(-)malic acid was transformed into 6, α _D +10.3° (c 0.91, CHCl₃) m.p. 49-50°C, R_F 0.50 in *n*hexane/EtOAc, 4/1 by vol, in 77% overall yield in 3 steps: (i) NaOMe in MeOH/THF, 1/1 by vol, (ii) t-BuPh₂SiCI-imidazole in dimethylformamide, (iii) NaOH in MeOH/THF, 1/1 by vol. In a similar way, (R)-2-t-butyldiphenylsilyloxytetracosanic acid 5, α _D -11.6^o $(c 0.29, CHCl₃), m.p. 48-49°C$, could be prepared in 50% overall yield from 8 [the $(-)$ -MTPA ester showed that 8 was obtained in 96.2%ee] in 4 steps *(via* 11, 12 and 13): *(i)* Ph₃P-PhCOOH-diethyl azodicarboxylate in THF [8], (ii) NaOMe in MeOH/THF, 1/1 by vol, (iii) *t-BuPh2SiCI/4-dimethylaminopyridine* in pyridine, (iv) NaOH in MeOH/THF, 1/1 by vol.

400 MHz ¹H-NMR of the (-)MTPA esters [9], (S)-10 and (R)-14, which were obtainable, respectively, from 6 and 5 in 3 steps; (i) CH_2N_2 , (ii) Bu_4NF in THF, (iii) (-)-MTPA chloride in pyridine, showed that (S) -10 and (R) -14 were obtained in 97.0% ee and 95.4% ee, respectively, and that no significant racemization occurred during these transformations.

A solution of 4 and (R)-acid 5 in CH_2Cl_2 was treated with 1,3-dicyclohexylcarbodiimide in the presence of hydroxy benzotriazole to give a 92% yield of completely protected $(2S)$ 3R, 4E, 2'R)-ceramide 15, R_F 0.52 and 0.45 in n-hexane/EtOAc, 4/1 by vol, which was solvolysed in MeOH/CH₂Cl₂, 1/1 by vol, in the presence of Amberlist[®] 15 to afford a 67% yield of the desired product 17, $\alpha|_D$ +6.4 α (c 1.13, CHCl₃), m.p. 44-45 α °C, R_F 0.30 in *n*-hexane/ EtOAc, 7/3 by vol. Treatment of 17 with Bu₄NF in THF afforded (2S, 3R, 4E, 2'R)-ceramide **19,** $[\alpha]_D + 8.7^{\circ}$ (c 1.13, CHCI₃/MeOH, 9/1 by vol), m.p 98-99 $^{\circ}$ C, R_F 0.45 in CHCI₃/MeOH, 9/1 by vol.

In a similar manner, 4 was condensed with (S) -acid 6 to give a 90% yield of completely protected $(2S, 3R, 4E, 2'S)$ -ceramide 16, R_F 0.62 and 0.55 in n-hexane/EtOAc, 4/1 by vol, which was further transformed into the oily glycosyl acceptor **18** (62%), α _D -6.4° (c 1.91, CHCI₃), R_F 0.46 in *n*-hexane/EtOAc, 7/3 by vol. Desilylation of **18** with fluoride anion gave an 80% yield of (2S, 3R, 4E, 2'S)-ceramide **20**, $[\alpha]_D$ -11.1° (c 1.41, CHCI₃/MeOH, 9/1 by vol), m.p. 100-101°C, R_F 0.51 in CHCl₃/MeOH, 9/1 by vol.

Crucial glycosylation of (2'R)-ceramide 17with galactopyranosyl trichloroacetimidate 3 in CHCI₃ in the presence of BF₃-ether and molecular sieves 4\AA according to the method of Schmidt and Michel $[5]$ afforded a 33% yield of the protected $(2'R)$ -cerebroside 21, $\alpha|_{\mathbf{D}}$ +1.6° (c 1.12, CHCl₃), R_F 0.45 in CHCl₃/MeOH, 49/1 by vol. Complete deprotection of 21, by (i) Bu₄NF in THF, (ii) NaOMe-MeOH, afforded a 62% yield of $(2'R)$ -cerebroside 1, $\alpha_{\rm lb}$ +8.5° (c 0.40, CHCI₃/MeOH, 1/1 by vol), R_F 0.37 in CHCI₃/MeOH, 17/3 by vol. Similarly, (2'S)-ceramide 18 was transformed into the protected cerebroside 22 (31%), $[\alpha]_D$ -2.8° (c 1.09, CHCl₃), R_F 0.61 in CHCl₃/MeOH, 49/1 by vol, which was further deprotected to afford (2'S)-cerebroside 2 (78%), α _D -25.2° (c 0.48, CHCI₃/MeOH, 1/1 by vol), R_F 0.48 in CHCI3/MeOH, 17/3 by vol.

In conclusion, (2'R) and (2'S)-cerebrosides 1 and 2 were synthesized in a stereo-controlled way and their 1 H-NMR data (Fig. 2) provided the conclusive evidence for the stereochemical assignment of the natural cerebroside.

Acknowledgements

We thank Dr. J. Uzawa and Mrs. T. Chijimatsu for recording and measuring the NMR spectra and Dr. H. Honma and his staff for the elemental analyses. We also thank Ms. A. Takahashi for her technical assistance.

References

- 1 Sugimoto M, Horisaki T, Ogawa T (1985) Glycoconjugate J 2:1145.
- 2 Hakomori S (1983) in Handbook of Lipid Research, Vol 3, Sphingolipid Biochemistry, eds. Kanfer JN, Hakomori S, Plenum Press, New York, p 1450.
- 3 Shapiro D, Flowers HM (1961) J Am Chem Soc 83:3327-32.
- 4 Dabrowski J, Egge H, Hanfland P (1980) Chem Phys Lipids 26:187-96.
- 5 Schmidt RR, Michel J (1980) Angew Chem Int Ed Engl 19:731-32.
- 6 Koike K, Nakahara Y, Ogawa T (1984) Glycoconjugate J 1:107-9.
- 7 Horn DHS, Pretorius YY (1954) J Chem Soc 1460-64.
- 8 Mitsunobu O, Eguchi M (1971) Bull Chem Soc Japan 44:3427-30.
- 9 Dale JA, Dull DL, Mosher HS (1969) J Org Chem 34:2543-49.